ANNUAL REVIEWS

Annual Review of Entomology

Spodoptera frugiperda: Ecology, Evolution, and Management Options of an Invasive Species

Wee Tek Tay,^{1,*} Robert L. Meagher Jr.,² Cecilia Czepak,³ and Astrid T. Groot⁴

¹CSIRO Black Mountain Laboratories, Australian Capital Territory, Australia; email: weetek.tay@csiro.au

²Agricultural Research Service, United States Department of Agriculture, Gainesville, Florida, USA; email: rob.meagher@usda.gov

³Escola de Agronomia, Campus Samambaia, Universidade Federal de Goiás, Goiânia, Brazil; email: ceciczepak@ufg.br

⁴Institute for Biodiversity and Ecosystem Dynamics, University of Amsterdam, Netherlands; email: a.t.groot@uva.nl

ANNUAL CONNECT

www.annualreviews.org

- Download figures
- · Navigate cited references
- · Keyword search
- Explore related articles
- Share via email or social media

Annu. Rev. Entomol. 2023. 68:299-317

First published as a Review in Advance on October 5, 2022

The *Annual Review of Entomology* is online at ento.annualreviews.org

https://doi.org/10.1146/annurev-ento-120220-102548

This work is licensed under a Creative Commons Attribution 4.0 International License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited. See credit lines of images or other third-party material in this article for license information.

*Corresponding author



Keywords

taxonomy, population genetics, host plant use, strain hybridization, mate attraction, insecticide resistance

Abstract

The fall armyworm (FAW), Spodoptera frugiperda (Lepidoptera, Noctuidae), is a well-known agricultural pest in its native range, North and South America, and has become a major invasive pest around the globe in the past decade. In this review, we provide an overview to update what is known about S. frugiperda in its native geographic ranges. This is followed by discussion of studies from the invaded areas to gain insights into S. frugiperda's ecology, specifically its reproductive biology, host plant use, status of insecticide resistance alleles, and biocontrol methods in native and invasive regions. We show that reference to host strains is uninformative in the invasive populations because multidirectional introduction events likely underpinned its recent rapid spread. Given that recent genomic analyses show that FAW is much more diverse than was previously assumed, and natural selection forces likely differ geographically, region-specific approaches will be needed to control this global pest.

1. INTRODUCTION

1.1. Taxonomy

Given the significant economic losses and food security challenges caused by the recent habitat expansion of the fall armyworm (FAW), *Spodoptera frugiperda*, it is important to clarify contradictory findings relating to this lepidopteran pest's strain, species, and hybrid status. As the corn budworm moth, *Phalaena frugiperda*, it was first described in 1797 from specimens collected in Georgia, United States (118). It was redescribed by de Boisduval & Guenée (22) as *Laphygma macra* in 1852, and synonymized with *Spodoptera* by Zimmerman in 1958 (151). From the 1840s through the 1860s, damaging populations in corn in Kansas, Missouri, and Illinois were very low in the spring but expanded in the late summer and fall. As a result of this distribution, which was explained as being a result of larvae or pupae overwintering in the soil, C.V. Riley (State Entomologist of Missouri), who did not know that the species had already been described, named it *Prodenia autumnalis*, the fall armyworm (109).

1.2. Detection of Host Strains in the Native Range

In 1985, two strains of FAW were identified and named after the host plants (i.e., corn and rice strains) from which they were collected (98). These strains are morphologically similar, although there is an indication that host plant habitat affects wing morphology (13, 81). Various genetic markers have been used to better understand their genetic variation and evolutionary relationship, including allozymes (96); amplified fragment length polymorphism (AFLP) (70, 106); polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) (66); the maternally inherited mitochondrial *cytochrome oxidase subunit I* (mtCOI) gene (64); the *Triosephosphate Isomerase* (*Tpi*) gene on the Z chromosome (82); simple sequence repeat (SSR) (i.e., microsatellite) loci (101); and more recently, genome-wide single-nucleotide polymorphism (SNP) markers and whole-genome resequencing (42, 108, 112, 124, 128, 150).

Soon after genetic differences were found between larvae collected from corn and those from bermudagrass or rice (96, 98), larval feeding studies were designed to test larval fitness from different plant hosts. While the first studies found developmental trend differences between the two strains on different host plants (133), further comparisons with larvae on bermudagrass and other grass species suggested more variable responses depending on season, artificial diets, and plants used (107).

Since host strains may also differ in their responses to constitutive plant defenses, the larvae's ability to metabolize the cyanide present in some pasture grasses (*Cynodon* spp.) (55) and salivary phospholipase C (PLC) activity levels (1) was also tested. Overall, only subtle differences between the two strains were identified in relation to host plant use, which has been confirmed by multiple studies (45, 58, 59). Therefore, these strains should not be called host strains, and we refer to them as C-strain and R-strain below. **Figure 1** outlines the basic biology of the FAW.

In addition to host plant differences, several prezygotic isolation barriers between the two strains have been investigated, including differential temporal patterns of female calling and copulation at night. In laboratory tests, the two strains were found to differ in the timing of female calling and copulation (51, 53, 97, 113, 114), with C-strain females being sexually active early at night and R-strain females late at night (but see 19, 110). Postzygotic isolation between the two strains has also been reported, with matings between C-strain females and R-strain males (CR matings) occurring significantly less than RC matings (27, 62, 110); hybrid RC females were found to be chaste, i.e., sexually completely inactive (62). Interestingly, 56–66% of all hybrids found in nature were shown to be RC hybrids (88, 90, 106). Thus, postzygotic isolation barriers seem to play a more important role than prezygotic isolation barriers (19, 110) and may explain

FAW: fall armyworm, the Entomological Society of America's official name for Spodoptera frugiperda (J. E. Smith)

AFLP: amplified fragment length polymorphism

PCR-RFLP: polymerase chain reaction-restriction fragment length polymorphism

mtCOI: maternally inherited mitochondrial (mt) cytochrome oxidase subunit I (COI) gene

Tpi: Triosephosphate Isomerase gene on the Z chromosome

SSR: simple sequence repeat

SNP:

single-nucleotide polymorphism

PLC: salivary phospholipase C; used by caterpillars against plant defense elicitors

C-strain: corn strain; original designation for the host strain found feeding on corn and other large grasses

R-strain: rice strain; original designation for the host strain found feeding on rice, pasture grasses, turf grasses, and other small grasses

Oviposition: egg clutches on plants

Egg clutches found on many plant species (1, 58, 136)



Egg clutches found in corn and sorghum fields, but also in other crops (19, 114, 122)

Two strains, C-strain and R-strain, with up to 2% genomic differences (50), although hybridization occurs between strains (10, 127)



No separate strains (127, 153)

No diapause, but adults migrate and remain active in (sub)tropical areas (70, 137).



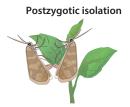
No diapause? Migration habits?

Sex pheromone components: Z9-14:OAc (major) (82, 132), Z7-12:OAc (2%) (21, 132, 133), Z11-16:OAc (132), Z9-12:OAc: function unclear) (133), E7-12:OAc (Brazil) (9)



Sex pheromone components: Z9-14:OAc (major), Z7-12:OAc (minor) (54, 64, 74)

Postzygotic isolation between C- and R-strain (28, 65)



Hybridization with, or postzygotic isolation, between locally occurring related *Spodoptera* species?

Figure 1

Basic biological differences in the life cycle of the fall armyworm between native and invasive populations.

the approximately 2% genomic divergence found between the two strains in their native range (26, 42). In addition to the C- and R-strains, Arias et al. (5) also reported two distinct C-strain clades in Brazil and Paraguay that diverged 0.65 Mya, which indicates that there are likely to be other genetically distinct native FAW populations.

1.3. Strain Hybridization in the Native Range

Whether assortative mating between the two strains occurs naturally across their native range is unclear (97, 99, 114). When the two strains were first identified, Prowell et al. (106) noted that as many as 16% of the collected individuals were hybrids, based on composite genotypes with mitochondrial DNA of one strain and esterase genotypes of the other. In laboratory choice tests,

analyses of spermatophores also demonstrated common occurrences of interstrain matings where spermatophore mtCOI markers were different from the mtCOI markers of the females (72). Strain and hybrid identifications over the past two decades were predominantly based on two markers, the mtCOI and the *Tpi* locus on the *Z* chromosome. However, since lepidopteran females are the heterogametic sex and possess only a single copy of the *Z* chromosome, a hybrid female or her offspring cannot be distinguished from a homozygous (i.e., nonhybrid) individual based on the *Tpi* marker or the nonrecombining mitochondrial genome (59). Analysis of whole-genome sequence data therefore offers a clearer picture of the level of hybridization between the two strains, and hybrids have now been identified from South America (5, 59, 124), Central America, the Caribbean, and North America (47, 112, 124).

1.4. Population Genetics in the Native Range

By 1913, it was known that the FAW had continuous generations in the tropics and subtropics (24). Later research suggested that outbreaks in the United States originated in Mexico and the West Indies and that the FAW overwinters in southern Texas; in southern Florida; and, in very warm winters, in the southern part of the Gulf Coast states (67, 134).

Genetic diversity of populations based on AFLP markers indicated significant gene flow, represented by high within-population variation and low between-population variation among North and South American populations (16, 69). However, local-scale isolation-by-distance between populations was detected in various South American populations (5, 9, 59). This highlights the difficulty of drawing general conclusions related to the FAW's propensity for long-distance migration, in which local and regional ecoclimatic factors are likely to also play important roles.

Seasonal migratory patterns of moths were inferred through haplotype ratios present in progenies of migrating populations (89) and through modeling-based genetic analysis (138, 139). These analyses showed that populations from Mexico, South America, and Trinidad and Tobago carried a haplotype ratio similar to that of moths from Texas (84, 91), while populations from Puerto Rico, the Dominican Republic, St. Kitts and Nevis, Dominica, and Barbados had haplotype ratios more similar to those of moths from Florida (84). These results thus suggest limited genetic exchange between Florida and South American FAW populations, as is also reported between distantly separated populations based on SSR markers (5, 101), although potential migrants were detected (101). However, lepidopteran SSR markers can be related to transposable elements (123) and could lead to misinterpretation of limited gene flow.

Based on 870 neutral and unlinked SNP markers throughout the genome, a principal component analysis grouped FAW populations from North America (Mississippi, Florida, Mexico), the Caribbean (Puerto Rico, Guadeloupe), and South America (French Guiana, Peru, Brazil) into five distinct populations that largely reflected their geographic regions (124) but not their strain identity, as was also reported by Schlum et al. (112). In line with the haplotype ratio findings (84, 91), Tay et al. (124) also showed that Guadeloupe and Puerto Rico FAW grouped with the Florida population, but separately from the Mississippi population. Furthermore, limited gene flow between North and South American populations (5, 9, 59) was detected, with one of the two Brazilian populations clustering with the Peruvian population (46, 143). In contrast, whole-genome SNP analysis (112) showed high gene flow between Caribbean and North and South American populations but also detected unique population clusters between Florida and Brazilian populations that suggested varying degrees of gene flow between the two continents.

2. INVASIONS OF FALL ARMYWORM

Invasion of the FAW into the Eastern Hemisphere and its resulting spread have been explained by two competing hypotheses, one positing a west-to-east spread and the other a complex of multiple

introductions involving African and Asian origins. There are some indications that, since the mid1960s, the FAW may have invaded the Eastern Hemisphere with increasing frequency. For example, adults that developed from larvae collected from crops in Israel in 1967 were identified as FAW
(140), although trapping in the Jordan Valley in 1979 with a FAW pheromone lure did not capture
any moths (145). In Germany, FAW-infected maize plants were collected and destroyed in August
1999 (57). In 2008, the FAW was reported to damage turfgrasses in parks surrounding Hanoi,
Vietnam (136). Prior to February 2014, morphological identification by the USDA Identification
Technology Program (ITP) of FAW-intercepted larvae entering US ports indicated that individuals were not only from native regions but also from countries such as Turkey, China, Indonesia,
Israel, Micronesia, the Netherlands, and Thailand (39). However, as far as we know, in none of
these cases are specimens available for molecular diagnostics or morphological reassessment.

The FAW was officially reported in western Africa in early 2016 (41), throughout the sub-Saharan African nations by early 2018 (29, 32), and in the Middle East/Indian subcontinent by mid-2018 (36). Single-gene analysis of light-trapped specimens suggested that the FAW had spread from Myanmar into southern China's Yunnan Province by early December 2018 (122), with ensuing northward expansion into central China (141). Eastward expansion would lead to its detection in South Korea and Japan, and southward expansion through southeast Asia allowed it to reach Australia by early 2020 (92, 108, 125). Chronological reports showing it moving from Africa progressively across to Asia, together with population genetic analysis based on single markers, therefore suggested a rapid west-to-east spread of the FAW (29, 32).

2.1. Origins of Invasive Populations

The widely accepted west-to-east spread hypothesis of invasive FAW across the Old World was inferred from strain-specific partial single gene markers (85). Specifically, the invasion analysis based on the Z-linked *Tpi* marker identified predominantly C-strain moths, originating either from the eastern United States or the Greater Antilles (87). This finding concurred with findings from West Africa, where the invasive descendants of C-strain and hybrid genotypes contained a pheromone composition similar to that found in the United States (51). Further single-marker analysis of moths from South Africa, India, Myanmar, and southern China indicated genetic backgrounds similar to those from west Africa (83, 86). Genomic analysis that included FAW from Louisiana, South Africa, Ethiopia, and Kenya (47) was also used to support the idea that China's Yunnan and Guangxi populations originated from Africa, despite having a genomic signature that indicated multiple independent introductions within China and in Africa.

The complex multiple introductions hypothesis involving African and Asian origins is supported by other recent multigenetic and genomic analyses, which indicate multiple introductions (95, 108, 112, 124, 125), as well as by the east-to-west movements of the pest (124). For example, genome-wide SNP markers clustered FAW from Benin, some individuals from Uganda, and populations from India and Yunnan China together but excluded Malawian and various Ugandan individuals (46, 94, 95, 124), which had a southeast Asian origin (108) (**Figure 2**). Similarly, whole-genome sequence analysis of Kenyan (112) and Zambian (150) FAW also supported multiple introductions. Phylogenetic analysis showed that African populations were predominantly derived from distinct Chinese populations, while some Chinese individuals were closely related to US populations (47, 124).

Similarly, unique genomic signatures shared between Yunnan and southeast Asian FAW populations (108) do not support the origin of Yunnan population being Myanmar or long-distance movements via monsoon winds (149), but instead support the hypothesis of multiple introductions in southeast Asia (**Figure 2**). Resistance allele characterization (11, 46, 49, 68) and different insecticide responses between Queensland and western Australian FAW populations also supported

ITP: United States Department of Agriculture Identification Technology Program

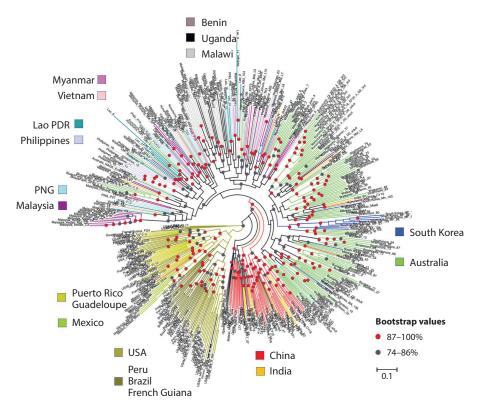


Figure 2

Maximum likelihood phylogeny of invasive (China, India, Australia, South Korea, Malaysia, Papua New Guinea, Philippines, Laos, Vietnam, Myanmar, Benin, Uganda, Malawi) and native [Puerto Rico, Guadeloupe, Mexico, United States (Florida, Mississippi), Peru, Brazil, French Guiana] populations. Fall armyworm populations are based on 870 neutral genome-wide single-nucleotide polymorphisms. Countries are indicated by branch colors. The derived state of African populations from southeast Asia is contrary to the axiom of west-to-east spread pattern. Evidence of multiple introductions can be seen from various populations, e.g., in Australia and Malaysia. Figure adapted with permission from Reference 108; CC BY-NC 4.0. Abbreviations: PDR, People's Democratic Republic; PNG, Papua New Guinea.

multiple non-African introductions into China, Indonesia, and northern Australia (108, 125). Understanding the degree of bidirectional gene flow between African, Asian, and Oceanian FAW, as well as genome diversity in countries such as Thailand, Cambodia, Indonesia, and many African and Asian countries, will require whole-genome analysis. Several of these countries may have had FAW introductions prior to 2016 (39).

2.2. Strain Differentiation in Invasive Populations?

Whether the two FAW strains have invaded other parts of the world has remained unclear. The confusion is in part due to disagreements between diagnostic markers, while usage of the terms strains, host-races, host forms, biotypes, and sibling and sister species to describe molecular diagnostics of the FAW has contributed to the confusion. Genetic analysis of Ugandan specimens documents both strains, with a higher genetic diversity in the C-strain than was previously reported (95). This higher genetic diversity in the C-strain in eastern Africa (e.g., Uganda, Malawi)

reflected a southeast Asian–derived origin: FAW populations from Vietnam, Myanmar, Malaysia, and Laos have greater frequencies and diversity of C-strain mitochondrial genomes that are absent in a majority of Chinese FAW (47, 124, 150). Analyses based on higher density of nuclear markers and whole-genome analyses in southeast Asian FAW further showed a complex mixing of genomes when the strain types are considered (47, 126, 150). Interestingly, while whole-genome resequencing and genome-wide SNP analyses identified the majority of African, Asian, and Australian invasive populations as hybrids, some Chinese (47, 126), Malaysian (Kedah State) (47, 124, 150), and African (47) individuals were identified as having non-admixed genomes. Global invasive FAW populations thus consist predominantly of hybridized individuals but also contain non-admixed individuals, both of which should be identified via a whole-genome analysis approach. The binary thinking that there are two distinguishable FAW strains throughout the invasive range should thus be avoided.

3. ECOLOGY

3.1. Mate Attraction in the Native Range

In addition to the host plant preference and performance studies, mating compatibility studies, and studies on variation in circadian rhythms mentioned in Section 1.2, variation in FAW mate attraction has been investigated throughout its native range. Mate attraction in the FAW occurs through a species-specific sex pheromone, which was identified in 1986 (129). The four main components are (*Z*)-9-tetradecenyl acetate (*Z*9–14:OAc), the main component; (*Z*)-7-dodecenyl acetate (*Z*7–12:OAc), the critical secondary sex pheromone component; and (*Z*)-11-hexadecenyl acetate (*Z*11–16:OAc) and *Z*9–12:OAc as minor components (see **Figure 1**). These components are used as lures in field trapping (79, 129, 130). In Brazil, an additional component, (*E*)-7-dodecenyl acetate (*E*7–12:OAc), was identified that increased male trapping in field tests (8).

Strain differences in female pheromone blends and attraction of males to females of different strains have been tested in several populations across the Americas. In field tests in Louisiana, 60% of males responded to same-strain females (97). However, in field tests in Florida, 68% of C-strain males responded to C-strain females, but 58% of R-strain males also responded to C-strain females (73). Identification of chemicals from the pheromone glands of the two strains in Colombia found nine compounds but no strain differences (12). Two other studies showed small differences in pheromone components between strains (44, 65). Follow-up field studies with lures with different blends showed that, in corn fields, more males of both strains responded to the C-strain blend than to the R-strain blend, while males of both strains in grass fields throughout North and South America were equally attracted to both blends (130, 131). Thus, differences in attraction based on pheromone blend between strains do not seem strong enough to cause assortative mating, although other factors such as differences in timing or means of pheromone emission might still cause assortative mating

3.2. Ecology in the Invasive Range

The ecology of FAW in the invasive ranges has been little investigated, as the focus of research has mostly been on trying to manage and control the pest. In this section, we summarize the ecological studies conducted with invasive FAW populations in relation to mate attraction, host use, and migration capabilities.

3.2.1. Mate attraction. Although many males were collected in commercial lure traps in West Africa (61, 71, 127), commercial FAW pheromone lures appear to be suboptimal in many invasive regions. This suggests that changes have occurred in the sexual communication of the FAW, either

Z9-14:OAc:

(Z)-9-tetradecenyl acetate, the main component of the FAW sex pheromone

Z7-12:OAc:

(Z)-7-dodecenyl acetate, a critical secondary component of the FAW sex pheromone

Z11-16:OAc:

(Z)-11-hexadecenyl acetate, a minor component of the FAW sex pheromone

Z9-12:OAc:

(Z)-9-dodecenyl acetate, a minor component of the FAW sex pheromone

E7-12:OAc:

(E)-7-dodecenyl acetate, a minor component found in the FAW sex pheromone in South America in the female sex pheromone or in the male response, possibly due to the predominantly hybrid nature of these populations. To date, only one study has been published on sex pheromones of moths; this studied moths from Africa and analyzed females collected as larvae from Benin and Nigeria (51). No significant differences were found between these populations and the Florida R-strain. However, male antennal responses showed significant differences, suggesting that males from different regions might respond differently.

3.2.2. Host plant use. In its invasive ranges, the FAW's use of host plants has been investigated to some extent (**Figure 1**). While maize has been the most widely damaged economic crop, other plants can also be attacked. For example, in Yangling, Shaanxi, China, FAW larvae were reported to develop fastest on maize but also developed well on other cash crops, especially wheat and, to a lesser extent, soybean, tomato, and cotton (137). On Chinese cabbage, the egg-hatching rate was low, with only 5.3% of larvae developing into egg-laying adults. Regardless, FAW larvae still fed voraciously on this plant and may use it as supplementary food during migration. In maize fields, competition may occur between the FAW and local pest species. For example, in laboratory feeding assays conducted in Yunnan, China, FAW larvae preyed on and outcompeted *Spodoptera litura* larvae on maize (119).

Host plant preference experiments have also been conducted in the FAW's invasive range, mostly to investigate the potential use of push-pull and intercropping management strategies. For example, testing oviposition preference on six grass species to determine the species' suitability as trap or pull plants showed that oviposition preference depended on plant size, although larvae mostly preferred maize (15). However, in no-choice oviposition tests, moths laid eggs on the grasses, indicating their potential usefulness as pull plants (15). As a repellent or push plant, Desmodium intortum was reported to significantly reduce numbers of FAW larvae on maize plants in Kenya, Uganda, and Tanzania (74). However, oviposition choice tests conducted in Kenya (111) also found that FAW laid similar number of eggs on D. intortum as on maize plants, and larvae could develop on D. intortum plants. In this region, Brachiara grasses were suggested as possible pull plants because FAW females oviposited similarly on these plants as on maize plants (111). Examples of intercropping experiments in Congo's Kashusha district also reported less frequent FAW attack on onion when intercropped with groundnut (17). Importantly, a population collected from maize fields in Zhejiang, China and established in the laboratory showed significant host plant adaptation (52). After 20 generations of selection on rice plants, larvae showed increased fitness (i.e., higher larval and pupal weight) and higher egg-laying rates on a rice plant host. This result suggests the potential for this pest to further impact agricultural productivity in China, as well as other Asian countries where rice is cultivated.

3.2.3. Migration capabilities. As mentioned above, the FAW is capable of long-distance migration. Population genomic studies of invasive populations suggest that long-distance movements occur frequently, either through natural migration and/or through anthropogenic activities (29, 32; see Section 2). In flight mill experiments, adult FAW developed from maize field-collected larvae in Yunnan, China could fly up to 120 km over five consecutive nights, with superior flight performance at 20–25°C and 60–90% relative humidity (38).

4. INSECTICIDE RESISTANCE IN FALL ARMYWORM

4.1. Insecticide Resistance in the Native Range

In native FAW populations, resistance to pyrethroids, carbamates, and organophosphates has been detected (14, 102); resistance to carbamates has been reported from Georgia, United States since

the 1970s (146), and resistance to pyrethroids was reported in 1997 in a laboratory-maintained population in Brazil (25). More recently, resistance to novel classes of insecticidal compounds [i.e., spinosad, *Bacillus thuringiensis* (Bt) toxins] was also reported. For example, spinosad resistance was detected in Brazil (93) and Puerto Rico (50), while resistance to the Bt Cry1F toxin expressed in transgenic maize was also reported in Puerto Rico (121) and Brazil (31). In addition, resistance to diamides in Brazil (10) and the VIP3A toxin in Louisiana FAW populations (144) has been reported.

Insecticides have been used to manage FAW populations since the early 1900s in the United States (67), which has led to the development of resistance to many insecticides. Monitoring for insecticide resistance frequency and response changes requires knowledge of baseline susceptibility levels. However, due to the long-term, early, and widespread use of insecticides on the FAW, baseline resistance levels are often unknown in target populations. Therefore, it is difficult to interpret how resistance levels have changed or evolved over time. The reported susceptible native FAW populations from North and South America (63, 147) represent valuable resources for monitoring insecticide response changes and for identification of the genetic basis underlying this resistance.

4.2. Insecticide Resistance in the Invasive Range

Research groups in Africa (46, 117), India (23, 63), China (46, 150), southeast Asia (11, 108), and Australia (92, 108, 125) have been investigating resistance status to various classes of insecticides in invasive FAW populations through bioassays, direct gene characterization via PCR and Sanger sequencing, and whole-genome resequencing. Unfortunately, the lack of consensus in bioassay approaches (e.g., chemical exposure methodologies, mortality rate scoring, insect life stages to be used, calculation of lethal dose) hinders meaningful comparisons between studies (23, 63).

Recently, insecticide resistance responses were compared between Indian invasive populations and a susceptible Brazilian native FAW population at spatial and temporal scales (63). These comparisons showed that responses to a range of insecticides have remained relatively constant between 2008 and 2019 and 2020. Changes in tolerance levels to some insecticides (e.g., chlorpyrifos and emamectin benzoate) were detected in some populations, although no genomic analysis was performed to rule out novel introduction events. In comparing differences in the response to methomyl among South African, Australian, and either susceptible or resistant Florida FAW populations (63, 147), Australian populations exhibited toxicity ratios within natural variability range. whereas high toxicity ratios were detected in the South African population (125). The characterization of resistance alleles supports the presence of unique FAW populations in southeast Asia, such as in Indonesia (11) and among different provinces in China (46, 68, 150). Notably, the same type of diamide resistance that was detected in Brazil (10) was also found in Guangzhou, China (68). Similarly, resistance to organophosphate and carbamate insecticides in native populations has been detected in populations from Hubei, China (49), but also in low frequency in Indonesia (11). These results indicate separate and potentially ongoing incursions of distinct New World FAW into the Old World.

Various FAW invasive populations exhibit unique resistance traits, and adoption of resistance management strategies developed from research on the assumed western African invasive bridgehead population (48) can lead to suboptimal outcomes. Understanding FAW introduction pathways and frequencies will help in monitoring future accidental introductions of novel resistance alleles and the ongoing management of this invasive pest, as well as in preparing for reciprocal introductions of undesirable genetic traits from the invasive range back to the native range.

Bt: Bacillus thuringiensis toxins, including Cry1F and VIP3a, that have been genetically inserted into crop plants

5. OTHER MANAGEMENT STRATEGIES

5.1. Use of Sex Pheromone Lures in the Native Range

SfNPV: *S. frugiperda* nucleopolyhedrovirus; used in microbial control of the FAW

In its native range, pest management strategies against the FAW include the use of sex pheromones as lures for monitoring. Sex pheromones have also been used as mating disruptants (75, 120), whereby the air is permeated with pheromone point sources to make it difficult for males to locate females, either due to confusion or because communication is disrupted as a result of habituation on the part of the perceiving insect. Release of Z9–12:OAc in an area where virgin females were calling reduced male captures by 85% (77). Subsequent studies showed that release of (Z,E)-9,12-tetradecadienyl acetate, a chemical that is not part of the FAW's pheromone, reduced the number of mated females by 87% (76). Expanded field studies in corn fields showed that aerial application of Z9–14:OAc formulated in hollow fibers reduced matings by 86% and oviposition by 84% (78). Use of pheromones for mass trapping has also been suggested (75), but of all techniques that include pheromone lures, this technique has the lowest chance of success for the FAW because of its high dispersal rate, multiple matings, and outbreak populations (7).

5.2. Use of Sex Pheromone Lures in the Invasive Range

Sexual communication can quickly evolve in novel environments to minimize interactions and communication interference with endemic species (43). Even though the sex pheromone composition of populations in Benin and Nigeria did not seem to differ from that of the Florida R-strain, elevated antennal sensitivity toward Z7–12:OAc was found in African males (51). In Togo and Benin, locally designed traps were successfully used to trap male moths and could provide a less expensive resource for farmers (71, 127). In Zambia, pheromone traps with a yellow insecticide-treated screen were found to trap more FAW than those with a black insecticide-treated screen, although potential influencing factors such as cropping practices were not specifically investigated (40). These findings suggest a need for region-specific pheromone lures in the different invaded areas for timely FAW monitoring and management.

5.3. Biocontrol in the Native Range

The FAW in its native range is attacked by a large number of pathogens (bacteria, fungi, viruses), parasitoids (Diptera and Hymenoptera), and predators (Coleoptera, Dermaptera, Hemiptera, Hymenoptera) (6, 37, 80). Aside from the results of a few studies (54, 132), little is known about the impact of natural enemies in reducing populations in agricultural systems. For augmentative releases, egg parasitoids are the easiest of the natural enemies to rear in large numbers. In Brazil, a series of studies showed that release of the egg parasitoids *Telenomus remus* Nixon and *Trichogramma pretiosum* Riley has the potential to reduce field populations, and enough information has been gathered about the number of released individuals (34, 104), dispersal capacity (103), and costs (135) to allow the development of a *T. remus* release program. A release program of *T. pretiosum* in corn has also been developed using data on the number of FAW males caught in pheromone traps (33). The predatory earwig *Doru luteipes* Scudder is another natural enemy being considered for augmentative release in corn (100). The presence of at least one pair of *D. luteipes* per plant was enough to keep the FAW population under control and promote a 7% increase in corn production (18), although this approach relies on the cost effectiveness of rearing the predator.

Different types of viruses have also been identified and explored as potential biological control agents (56). The most promising viral candidates for biological control are the *S. frugiperda* nucleopolyhedrovirus (SfNPV) (105) and granuloviruses (20). The rerelease of the

nucleopolyhedrovirus in a cotton production program caused high mortality of caterpillars and reduced damage to the reproductive plant structures (21).

5.4. Biocontrol in the Invasive Range

Although the FAW is native to the New World, many natural enemy species that target it have already been documented across sub-Saharan Africa (3, 28, 60, 94, 116) and Asia (35, 115, 148). Mass rearing of egg parasitoids for inoculative release has commenced in both Africa (2, 126) and Nepal (30). In India, up to 73% of FAW larvae were parasitized by endemic parasitoid wasps or infected with the naturally occurring entomopathogenic fungus *Metarhizium rileyi* (35). Fungi such as *M. rileyi* and *Beauveria bassiana* therefore have promise as effective biocontrol agents to complement integrative pest management strategies, although their efficacy in causing mortality in the FAW appeared to be strain specific. The efficacy of intrahost specificity of SfNPV against invading host populations requires further characterization, including via screening of naturally occurring field isolates from China, India, and Nigeria (56).

6. FALL ARMY WORM EVOLUTIONARY POTENTIAL

Genomic analyses have shown that FAW populations in both native and invasive ranges are genetically much more diverse than was previously assumed. In its native range, strain-specific chemosensory and detoxification genes against plant metabolites and insecticides have been identified (42). Such gene expansions likely contribute to adaptation to novel habitats and to the detection, identification, and utilization of diverse plant hosts as food sources and oviposition sites, as well as assisting in long-distance flight (42, 142, 150). In invasive populations, new mutations, such as gene translocation (150) and composition differences of transposable elements in the genomes (142, 150), seem to occur relatively frequently across the FAW's genome. In addition to the high genetic diversity found in many FAW populations, human-assisted long-distance movements of the FAW can reciprocate introductions of genotypes from invasive populations to native New World populations.

Natural selection forces exerted on FAW individuals and populations will likely differ from region to region, as interacting species, including natural enemies, may vary locally. Sympatrically occurring species in the native range have most likely evolved mechanisms to prevent cross-attraction. However, in invasive areas, introgression and hybridization may have occurred between the introduced and native species, as reported in other noctuid pests, resulting in novel genetic variants (4). As mentioned above, extensive hybridization has already been shown to occur between FAW strains in native and invasive areas, although hybridization between FAW and other *Spodoptera* and noctuid species in the invaded regions has not yet been discovered. The introductions of the FAW to all (sub)tropical regions of the world came with significant ecological and socioeconomic costs, but studying the ecology and evolution of this species also creates unprecedented opportunities in the age of genomics to understand adaptation and evolutionary potentials under real-world conditions.

7. CONCLUSIONS

In reviewing the literature, including the most recent genomic analyses, we can draw six important conclusions. First, the FAW does not consist of two distinguishable host strains in the invaded areas. Second, the spread of invasive FAW across the Old World likely involved both west-to-east and east-to-west introduction events, although specimen confirmation of multiple introductions before 2016 is needed. Third, some commercial pheromone lures are not effective in several invasive regions, which may be due to regional variation in male responses. Fourth,

local host plant adaptation may occur relatively rapidly. Fifth, insecticide resistance responses are highly variable. Finally, biocontrol is likely to be most effective with endemic (egg) parasitoids, viruses, and entomopathogenic fungi.

DISCLOSURE STATEMENT

The authors are not aware of any affiliations, memberships, funding, or financial holdings that might be perceived as affecting the objectivity of this review.

ACKNOWLEDGMENTS

The use of trade, firm, or corporation names in this publication is for the information and convenience of the reader. Such use does not constitute an official endorsement or approval by the United States Department of Agriculture or the Agricultural Research Service of any product or service to the exclusion of others that may be suitable. W.T.T. was supported by CSIRO Health & Biosecurity, R.L.M. was supported by the United States Department of Agriculture, C.C. was supported by the Universidade Federal de Goiás, and A.T.G. was supported by the University of Amsterdam.

LITERATURE CITED

- Acevedo FE, Peiffer M, Ray S, Meagher R, Luthe DS, Felton GW. 2018. Intraspecific differences in plant defense induction by fall armyworm strains. New Phytol. 218:310–21
- Agboyi LK, Goergen G, Beseh P, Mensah SA, Clottey VA, et al. 2020. Parasitoid complex of fall armyworm, Spodoptera frugiperda, in Ghana and Benin. Insects 11:68
- Agboyi LK, Layodé BFR, Fening KO, Beseh P, Clottey VA, et al. 2021. Assessing the potential of inoculative field releases of *Telenomus remus* to control *Spodoptera frugiperda* in Ghana. *Insects* 12:665
- Anderson CJ, Oakeshott JG, Tay WT, Gordon KHJ, Zwick A, Walsh TK. 2018. Hybridization and gene flow in the mega-pest lineage of moth, Helicoverpa. PNAS 115:5034–39
- Arias O, Cordeiro E, Corrêa AS, Domingues FA, Guidolin AS, Omoto C. 2019. Population genetic structure and demographic history of Spodoptera frugiperda (Lepidoptera: Noctuidae): implications for insect resistance management programs. Pest Manag. Sci. 75:2948–57
- Ashley TR. 1979. Classification and distribution of fall armyworm parasites. Fla. Entomol. 62:114–23
- Barclay HJ. 1984. Pheromone trapping models for pest control: effects of mating patterns and immigration. Res. Popul. Ecol. 26:303–11
- Batista-Pereira LG, Stein K, de Paula AF, Moreira JA, Cruz I, et al. 2006. Isolation, identification, synthesis, and field evaluation of the sex pheromone of the Brazilian population of Spodoptera frugiperda. 7. Chem. Ecol. 32:1085–99
- Belay DK, Clark PL, Skoda SR, Isenhour DJ, Molina-Ochoa J, et al. 2012. Spatial genetic variation among Spodoptera frugiperda (Lepidoptera: Noctuidae) sampled from the United States, Puerto Rico, Panama, and Argentina. Ann. Entomol. Soc. Am. 105:359

 –67
- Boaventura D, Bolzan A, Padovez FEO, Okuma DM, Omoto C, Nauen R. 2020. Detection of a ryanodine receptor target-site mutation in diamide insecticide resistant fall armyworm, Spodoptera frugiperda. Pest Manag. Sci. 76:47–54
- Boaventura D, Martin M, Pozzebon A, Mota-Sanchez D, Nauen R. 2020. Monitoring of target-site mutations conferring insecticide resistance in Spodoptera frugiperda. Insects 11:545
- Cañas-Hoyos N, Lobo-Echeverri T, Saldamando-Benjumea CI. 2017. Chemical composition of female sexual glands of Spodoptera frugiperda corn and rice strains from Tolima, Colombia. Southwest. Entomol. 42:375–94
- Cañas-Hoyos N, Marquez EJ, Saldamando-Benjumea CI. 2014. Differentiation of Spodoptera frugiperda (Lepidoptera: Noctuidae) corn and rice strains from central Colombia: a wing morphometric approach. Ann. Entomol. Soc. Am. 107:575–81

- Carvalho RA, Omoto C, Field LM, Williamson MS, Bass C. 2013. Investigating the molecular mechanisms of organophosphate and pyrethroid resistance in the fall armyworm Spodoptera frugiperda. PLOS ONE 8(4):e62268
- Cheruiyot D, Morales XC, Chidawanyika F, Bruce TJA, Khan ZR. 2021. Potential roles of selected forage grasses in management of fall armyworm (*Spodoptera frugiperda*) through companion cropping. *Entomol. Exp. Appl.* 169:966–74
- Clark PL, Molina-Ochoa J, Martinelli S, Skoda SR, Isenhour DJ, et al. 2007. Population variation of the fall armyworm, Spodoptera frugiperda, in the Western Hemisphere. J. Insect Sci. 7:5
- Cokola MC, Ndjadi SS, Bisimwa EB, Ahoton LE, Francis F. 2021. First report of Spodoptera frugiperda (Lepidoptera: Noctuidae) on onion (Allium cepa L.) in South Kivu, Eastern DR Congo. Rev. Bras. Entomol. 65:e20200083
- Cruz I, Valicente FH. 1992. Manejo da lagarta-do-cartucho, Spodoptera frugiperda, em milho, usando o predador Doru luteipes e baculovirus. In Relatorio Tecnico Anual CNPMS 1988–1991, pp. 74–75. Sete Lagoas, Braz.: EMPRABPA
- Cruz-Esteban S, Rojas JC, Malo EA. 2017. Calling behavior, copulation time, and reproductive compatibility of corn-strain fall armyworm (Lepidoptera: Noctuidae) from populations in Mexico. *Environ. Entomol.* 46:901–6
- Cuartas P, Barrera G, Barreto E, Villamizar L. 2014. Characterisation of a Colombian granulovirus (Baculoviridae: Betabaculovirus) isolated from Spodoptera frugiperda (Lepidoptera: Noctuidae) larvae. Biocontrol Sci. Technol. 24:1265–85
- Czepak C, Godinho KCA, da Costa Gontijo P, Rezende JM. 2019. Cotton. In Natural Enemies of Insect Pests in Neotropical Agroecosystems, ed. B Souza, LL Vázquez, RC Marucci, pp. 293–303. Berlin: Springer
- de Boisduval JBAD, Guenée A. 1852. Histoire Naturelle des Insectes. Species General des Lepidopteres. Tome Cinquieme. Noctuelites. Tome 1. Paris: Roret
- Deshmukh S, Pavithra HB, Kalleshwaraswamy CM, Shivanna BK, Maruthi MS, Mota-Sanchez D. 2020.
 Field efficacy of insecticides for management of invasive fall armyworm, Spodoptera frugiperda (J. E. Smith) (Lepidoptera: Noctuidae) on maize in India. Fla. Entomol. 103:221–27
- 24. Dew JA. 1913. Fall army worm Laphygma frugiperda (S. & A.). 7. Econ. Entomol. 6:361–66
- Diez-Rodriguez GI, Omoto C. 2001. Inheritance of lambda-cyhalothrin resistance in Spodoptera frugiperda (J.E. Smith) (Lepidoptera: Noctuidae). Neotrop. Entomol. 30:311–16
- Dumas P, Barbut J, Le Ru B, Silvain JF, Clamens AL, et al. 2015. Phylogenetic molecular species delimitations unravel potential new species in the pest genus Spodoptera Gueneé, 1852 (Lepidoptera, Noctuidae). PLOS ONE 10:e0122407
- Dumas P, Legeai F, Lemaitre C, Scaon E, Orsucci M, et al. 2015. Spodoptera frugiperda (Lepidoptera: Noctuidae) host-plant variants: two host strains or two distinct species? Genetica 143:305–16
- Durocher-Granger L, Mfune T, Musesha M, Lowry A, Reynolds K, et al. 2021. Factors influencing the occurrence of fall armyworm parasitoids in Zambia. 7. Pest Sci. 94:1133–46
- Early R, González-Moreno P, Murphy ST, Day R. 2018. Forecasting the global extent of invasion of the cereal pest Spodoptera frugiperda, the fall armyworm. NeoBiota 40:25–50
- Elibariki N, Bajracharya ASR, Bhat B, Tefera T, Mottern JL, et al. 2020. Candidates for augmentative biological control of Spodoptera frugiperda in Kenya, Tanzania and Nepal. Indian J. Entomol. 82:606–9
- Farias JR, Andow DA, Horikoshi RJ, Sorgatto RJ, Fresia P, et al. 2014. Field-evolved resistance to Cry1F maize by Spodoptera frugiperda (Lepidoptera: Noctuidae) in Brazil. Crop Prot. 64:150–58
- Feldmann F, Rieckmann U, Winter S. 2019. The spread of the fall armyworm Spodoptera frugiperda in Africa—what should be done next? 7. Plant Dis. Prot. 126:97–101
- Figueiredo MLC, Cruz I, da Silva RB, Foster JE. 2015. Biological control with *Trichogramma pretiosum* increases organic maize productivity by 19.4%. *Agron. Sustain. Dev.* 35:1175–83
- 34. Figueiredo MLC, Della Lucia TMC, Cruz I. 2002. Effect of *Telenomus remus* Nixon (Hymenoptera: Scelionidae) density on control of *Spodoptera frugiperda* (Smith) (Lepidoptera: Noctuidae) egg masses upon release in a maize field. *Rev. Brus. Milho Sorgo* 1:12–19
- Firake DM, Behere GT. 2020. Natural mortality of invasive fall armyworm, Spodoptera frugiperda (J. E. Smith) (Lepidoptera: Noctuidae) in maize agroecosystems of northeast India. Biol. Control 148:104303

- Ganiger PC, Yeshwanth HM, Muralimohan K, Vinay N, Kumar ARV, Chandrashekara K. 2018. Occurrence of the new invasive pest, fall armyworm, Spodoptera frugiperda (J.E. Smith) (Lepidoptera: Noctuidae), in the maize fields of Karnataka, India. Curr. Sci. 115:621–23
- Gardner WA, Fuxa JR. 1980. Pathogens for the suppression of the fall armyworm. Fla. Entomol. 63:439–47
- Ge S-s, He L-m, He W, Yan R, Wyckhuys KAG, Wu K-m. 2021. Laboratory-based flight performance of the fall armyworm, Spodoptera frugiperda. J. Integr. Agric. 20:707–14
- Gilligan TM, Passoa SC. 2014. LepIntercept: an identification resource for intercepted Lepidoptera larvae. Identification Technology Program, USDA/APHIS/PPQ/S&T, Fort Collins, CO. https://idtools.org/ id/leps/lepintercept
- Gilson C, Francisco G, Bingham GV, Matimelo M. 2018. Efficacy of a pheromone trap with insecticide-treated long-lasting screen against fall armyworm (FAW), Spodoptera frugiperda (Lepidoptera: Noctuidae). Outlooks Pest Manag. 29:215–19
- Goergen G, Kumar PL, Sankung SB, Togola A, Tamò M. 2016. First report of outbreaks of the fall armyworm Spodoptera frugiperda (J E Smith) (Lepidoptera, Noctuidae), a new alien invasive pest in West and Central Africa. PLOS ONE 11:e0165632
- 42. Gouin A, Bretaudeau A, Nam K, Gimenez S, Aury J-M, et al. 2017. Two genomes of highly polyphagous lepidopteran pests (Spodoptera frugiperda, Noctuidae) with different host-plant ranges. Sci. Rep. 7:11816
- Groot AT, Dekker T, Heckel DG. 2016. The genetic basis of pheromone evolution in moths. Annu. Rev. Entomol. 61:99–117
- 44. Groot AT, Marr M, Schoff G, Lorenz S, Svatos A, Heckel DG. 2008. Host strain specific sex pheromone variation in *Spodoptera frugiperda*. Front. Zool. 5:20
- Groot AT, Unbehend M, Hänniger S, Juárez ML, Kost S, Heckel DG. 2016. Evolution of reproductive isolation of Spodoptera frugiperda. In Sexual Communication in Moths, ed. J Allison, R Cardé, pp. 291–300. Oakland, CA: Univ. Calif. Press
- Guan F, Zhang J, Shen H, Wang X, Padovan A, et al. 2021. Whole-genome sequencing to detect mutations associated with resistance to insecticides and Bt proteins in *Spodoptera frugiperda*. *Insect Sci.* 28:627–38
- Gui F, Lan T, Zhao Y, Guo W, Dong Y, et al. 2020. Genomic and transcriptomic analysis unveils
 population evolution and development of pesticide resistance in fall armyworm Spodoptera frugiperda.
 Protein Cell 13:513–31
- 48. Guillemaud T, Ciosi M, Lombaert E, Estoup A. 2011. Biological invasions in agricultural settings: insights from evolutionary biology and population genetics. C. R. Biol. 334:237–46
- Guo Z-M, Deng X-Q, Li J, Yuan M-J, Wan H, et al. 2020. Detection of insecticide sensitivity and target site mutations in field populations of *Spodoptera frugiperda* (Lepidoptera: Noctuidae) in four regions of Hubei, central China. *Acta Entomol. Sin.* 63:582–89
- Gutiérrez-Moreno R, Mota-Sanchez D, Blanco CA, Whalon ME, Terán-Santofimio H, et al. 2019.
 Field-evolved resistance of the fall armyworm (Lepidoptera: Noctuidae) to synthetic insecticides in Puerto Rico and Mexico. J. Econ. Entomol. 112:792–802
- Haenniger S, Goergen G, Akinbuluma MD, Kunert M, Heckel DG, Unbehend M. 2020. Sexual communication of Spodoptera frugiperda from West Africa: adaptation of an invasive species and implications for pest management. Sci. Rep. 10:2892
- Hafeez M, Li X, Ullah F, Zhang Z, Zhang J, et al. 2021. Behavioral and physiological plasticity provides insights into molecular based adaptation mechanism to strain shift in *Spodoptera frugiperda*. *Int. J. Mol.* Sci. 22:10284
- Hänniger S, Dumas P, Schöff G, Gebauer-Jung S, Vogel H, et al. 2017. Genetic basis of allochronic differentiation in the fall armyworm. BMC Evol. Biol. 17:68
- Harrison RD, Thierfelder C, Baudron F, Chinwada P, Midega C, et al. 2019. Agro-ecological options for fall armyworm (*Spodoptera frugiperda* JE Smith) management: providing low-cost, smallholder friendly solutions to an invasive pest. *J. Environ. Manag.* 243:318–30
- Hay-Roe MM, Meagher RL, Nagoshi RN. 2011. Effects of cyanogenic plants on fitness in two host strains of the fall armyworm (Spodoptera frugiperda). J. Chem. Ecol. 37:1314–22

- Hussain AG, Wennmann JT, Goergen G, Bryon A, Ros VID. 2021. Viruses of the fall armyworm Spodoptera frugiperda: a review with prospects for biological control. Viruses 13:2220
- Jeger M, Bragard C, Caffier D, Candresse T, Chatzivassiliou E, et al. 2017. Pest categorisation of Spodoptera frugiperda. EFSA J. 15(7):e04927
- Juárez ML, Murúa MG, Garcia MG, Ontivero M, Vera MT, et al. 2012. Host association of Spodoptera frugiperda (Lepidoptera: Noctuidae) corn and rice strains in Argentina, Brazil, and Paraguay. J. Econ. Entomol. 105:573–82
- Juárez ML, Schöfl G, Vera MT, Vilardi JC, Murúa MG, et al. 2014. Population structure of Spodoptera frugiperda maize and rice host forms in South America: Are they host strains? Entomol. Exp. Appl. 152:182–99
- 60. Kenis M, du Plessis H, Van den Berg J, Ba MN, Goergen G, et al. 2019. Telenomus remus, a candidate parasitoid for the biological control of Spodoptera frugiperda in Africa, is already present on the continent. Insects 10:10
- Koffi D, Agboka K, Adjevi AKM, Assogba K, Fening KO, et al. 2021. Trapping Spodoptera frugiperda (Lepidoptera: Noctuidae) moths in different crop habitats in Togo and Ghana. J. Econ. Entomol. 114:1138–44
- 62. Kost S, Heckel DG, Yoshido A, Marec F, Groot AT. 2016. A Z-linked sterility locus causes sexual abstinence in hybrid females and facilitates speciation in *Spodoptera frugiperda*. *Evolution* 70:1418–27
- Kulye M, Mehlhorn S, Boaventura D, Godley N, Venkatesh SK, et al. 2021. Baseline susceptibility of Spodoptera frugiperda populations collected in India towards different chemical classes of insecticides. Insects 12:758
- Levy HC, Garcia-Maruniak A, Maruniak JE. 2002. Strain identification of Spodoptera frugiperda (Lepidoptera: Noctuidae) insects and cell line: PCR-RFLP of cytochrome oxidase C subunit I gene. Fla. Entomol. 85:186–90
- Lima ER, McNeil JN. 2009. Female sex pheromones in the host races and hybrids of the fall armyworm, Spodoptera frugiperda (Lepidoptera: Noctuidae). Chemoecology 19:29–36
- Lu Y-J, Adang MJ, Isenhour DJ, Kochert GD. 1992. RFLP analysis of genetic variation in North American populations of the fall armyworm moth Spodoptera frugiperda (Lepidoptera: Noctuidae). Mol. Ecol. 1:199–207
- 67. Luginbill P. 1928. The fall army worm. Tech. Bull. 34, U. S. Dep. Agric., Washington, DC
- Lv D, Liu X, Dong Y, Yan Z, Zhang X, et al. 2021. Comparison of gut bacterial communities of fall armyworm (Spodoptera frugiperda) reared on different host plants. Int. J. Mol. Sci. 22:16
- Martinelli S, Clark PL, Zucchi MI, Silva-Filho MC, Foster JE, Omoto C. 2007. Genetic structure and molecular variability of *Spodoptera frugiperda* (Lepidoptera: Noctuidae) collected in maize and cotton fields in Brazil. *Bull. Entomol. Res.* 97:225–31
- McMichael M, Prowell DP. 1999. Differences in amplified fragment-length polymorphisms in fall armyworm (Lepidoptera: Noctuidae) host strains. Ann. Entomol. Soc. Am. 92:175–81
- Meagher RL, Agboka K, Tounou AK, Koffi D, Agbevohia KA, et al. 2019. Comparison of pheromone trap design and lures for Spodoptera frugiperda in Togo and genetic characterization of moths caught. Entomol. Exp. Appl. 167:507–16
- Meagher RL, Nagoshi RN. 2010. Identification of fall armyworm (Lepidoptera: Noctuidae) host strains based on male-derived spermatophores. Fla. Entomol. 93:191–97
- Meagher RL, Nagoshi RN, Armstrong JS, Niogret J, Epsky ND, Flanders KL. 2013. Captures and host strains of fall armyworm (Lepidoptera: Noctuidae) males in traps baited with different commercial pheromone blends. Fla. Entomol. 96:729

 –40
- Midega CAO, Pittchar JO, Pickett JA, Hailu GW, Khan ZR. 2018. A climate-adapted push-pull system
 effectively controls fall armyworm, Spodoptera frugiperda (J E Smith), in maize in East Africa. Crop Prot.
 105:10–15
- Mitchell ER. 1986. Pheromones: as the glamour and glitter fade—the real work begins. Fla. Entomol. 69:132–39
- Mitchell ER, Baumhover AH, Jacobson M. 1976. Reduction of mating potential of male *Heliothis* spp. and *Spodoptera frugiperda* in field plots treated with disruptants. *Environ. Entomol.* 5:484–86

- Mitchell ER, Copeland WW, Sparks AN, Sekul AA. 1974. Fall armyworm: disruption of pheromone communication with synthetic acetates. *Environ. Entomol.* 3:778–80
- 78. Mitchell ER, McLaughlin JR. 1982. Suppression of mating and oviposition by fall armyworm and mating by corn earworm in corn, using the air permeation technique. *7. Econ. Entomol.* 75:270–74
- Mitchell ER, Tumlinson JH, McNeil JN. 1985. Field evaluation of commercial pheromone formulations and traps using a more effective sex pheromone blend for the fall armyworm (Lepidoptera: Noctuidae). 7. Econ. Entomol. 78:1364–69
- Molina-Ochoa J, Carpenter JE, Heinrichs EA, Foster JE. 2003. Parasitoids and parasites of Spodoptera frugiperda (Lepidoptera: Noctuidae) in the Americas and Caribbean Basin: an inventory. Fla. Entomol. 86:254–89
- Nagoshi KL, Allan SA, Meagher RL. 2020. Assessing the use of wing morphometrics to identify fall armyworm (Lepidoptera: Noctuidae) host strains in field collections. *J. Econ. Entomol.* 113:800–7
- 82. Nagoshi RN. 2010. The fall armyworm triose phosphate isomerase (*Tpi*) gene as a marker of strain identity and interstrain mating. *Ann. Entomol. Soc. Am.* 103:283–92
- Nagoshi RN, Dhanani I, Asokan R, Mahadevaswamy HM, Kalleshwaraswamy CM, et al. 2019. Genetic characterization of fall armyworm infesting South Africa and India indicate recent introduction from a common source population. PLOS ONE 14:e0236759
- 84. Nagoshi RN, Fleischer S, Meagher RL, Hay-Roe M, Khan A, et al. 2017. Fall armyworm migration across the Lesser Antilles and the potential for genetic exchanges between North and South American populations. PLOS ONE 12:e0171743
- Nagoshi RN, Goergen G, Du Plessis H, van den Berg J, Meagher R. 2019. Genetic comparisons of fall armyworm populations from 11 countries spanning sub-Saharan Africa provide insights into strain composition and migratory behaviors. Sci. Rep. 9:8311
- Nagoshi RN, Htain NN, Boughton D, Zhang L, Xiao YT, et al. 2020. Southeastern Asia fall armyworms are closely related to populations in Africa and India, consistent with common origin and recent migration. Sci. Rep. 10:10
- Nagoshi RN, Koffi D, Agboka K, Tounou KA, Banerjee R, et al. 2017. Comparative molecular analyses
 of invasive fall armyworm in Togo reveal strong similarities to populations from the eastern United
 States and the Greater Antilles. PLOS ONE 12:e0181982
- Nagoshi RN, Meagher R. 2003. Fall armyworm FR sequences map to sex chromosomes and their distribution in the wild indicate limitations in interstrain mating. Insect Mol. Biol. 12:453–58
- Nagoshi RN, Meagher RL, Flanders K, Gore J, Jackson R, et al. 2008. Using haplotypes to monitor the migration of fall armyworm (Lepidoptera: Noctuidae) corn-strain populations from Texas and Florida. 7. Econ. Entomol. 101:742–49
- Nagoshi RN, Meagher RL, Nuessly G, Hall DG. 2006. Effects of fall armyworm (Lepidoptera: Noctuidae) interstrain mating in wild populations. *Environ. Entomol.* 35:561–68
- Nagoshi RN, Rosas-García NM, Meagher RL, Fleischer SJ, Westbrook JK, et al. 2015. Haplotype profile comparisons between Spodoptera frugiperda (Lepidoptera: Noctuidae) populations from Mexico with those from Puerto Rico, South America, and the United States and their implications to migratory behavior. J. Econ. Entomol. 108:135–44
- Nguyen DT, Chen Y, Herron GA. 2021. Preliminary characterisation of known pesticide resistance alleles in Spodoptera frugiperda (Lepidoptera: Noctuidae) in its invasive Australian range. Austral Entomol. 60:782–90
- Okuma DM, Bernardi D, Horikoshi RJ, Bernardi O, Silva AP, Omoto C. 2017. Inheritance and fitness costs of Spodoptera frugiperda (Lepidoptera: Noctuidae) resistance to spinosad in Brazil. Pest Manag. Sci. 74:1441–48
- Otim MH, Aropet SA, Opio M, Kanyesigye D, Opolot HN, Tay WT. 2021. Parasitism of fall armyworm Spodoptera frugiperda (Lepidoptera: Noctuidae) in different maize producing regions of Uganda. Insects 12:121
- Otim MH, Tay WT, Walsh TK, Kanyesigye D, Adumo S, et al. 2018. Detection of sister-species in invasive populations of the fall armyworm Spodoptera frugiperda (Lepidoptera: Noctuidae) from Uganda. PLOS ONE 13:18

- Pashley DP. 1986. Host-associated genetic differentiation in fall armyworm (Lepidoptera: Noctuidae): a sibling species complex? Ann. Entomol. Soc. Am. 79:898–904
- Pashley DP, Hammond AM, Hardy TN. 1992. Reproductive isolating mechanisms in fall armyworm host strains (Lepidoptera: Noctuidae). Ann. Entomol. Soc. Am. 85:400–5
- Pashley DP, Johnson SJ, Sparks AN. 1985. Genetic population structure of migratory moths: the fall armyworm (Lepidoptera: Noctuidae). Ann. Entomol. Soc. Am. 78:756–62
- Pashley DP, Martin JA. 1987. Reproductive incompatibility between host strains of the fall armyworm (Lepidoptera: Noctuidae). Ann. Entomol. Soc. Am. 80:731–33
- Pasini A, Parra JRP, Lopes JM. 2007. Artificial diet for rearing *Doru luteipes* (Scudder) (Dermaptera: For-ficulidae), a predator of the fall armyworm, *Spodoptera frugiperda* (J.E. Smith) (Lepidoptera: Noctuidae).
 Neotrop. Entomol. 36:308–11
- Pavinato VAC, Martinelli S, de Lima PF, Zucchi MI, Omoto C. 2013. Microsatellite markers for genetic studies of the fall armyworm, Spodoptera frugiperda. Genet. Mol. Res. 12:370–80
- Pitre HN. 1988. Relationship of fall armyworm (Lepidoptera: Noctuidae) from Florida, Honduras,
 Jamaica, and Mississippi: susceptibility to insecticides with reference to migration. Fla. Entomol. 71:56–61
- 103. Pomari-Fernandes A, de Freitas Bueno A, De Bortoli SA, Favetti BM. 2018. Dispersal capacity of the egg parasitoid *Telenomus remus* Nixon (Hymenoptera: Platygastridae) in maize and soybean crops. *Biol. Control* 126:158–68
- 104. Pomari-Fernandes A, de Freitas Bueno A, Oliveira de Freitas Bueno RC, Menezes A Jr., Prado Fernandes Fonseca AC. 2013. Releasing number of *Telenomus remus* (Nixon) (Hymenoptera: Platygastridae) against *Spodoptera frugiperda* Smith (Lepidoptera: Noctuidae) in corn, cotton and soybean. *Cienc. Rural* 43:377– 82
- 105. Popham HJR, Rowley DL, Harrison RL. 2021. Differential insecticidal properties of Spodoptera frugiperda multiple nucleopolyhedrovirus isolates against corn-strain and rice-strain fall armyworm, and genomic analysis of three isolates. 7. Invertebr. Pathol. 183:107561
- Prowell DP, McMichael M, Silvain JF. 2004. Multilocus genetic analysis of host use, introgression, and speciation in host strains of fall armyworm (Lepidoptera: Noctuidae). Ann. Entomol. Soc. Am. 97:1034

 –44
- Quisenberry SS, Whitford F. 1988. Evaluation of bermudagrass resistance to fall armyworm (Lepidoptera: Noctuidae): influence of host strain and dietary conditioning. 7. Econ. Entomol. 81:1463–68
- Rane R, Walsh TK, Lenancker P, Gock A, Dao TH, et al. 2022. Complex multiple introductions drive fall armyworm invasions into Asia and Australia. bioRxiv 2022.06.11.495773. https://doi.org/10.1101/ 2022.06.11.495773
- 109. Riley CV. 1870. The fall armyworm. Prodenia autumnalis, n. sp. Am. Entomol. Bot. 2:363-65
- Saldamando-Benjumea CI, Estrada-Piedrahíta K, Velásquez-Vélez MI, Bailey RI. 2014. Assortative mating and lack of temporality between corn and rice strains of Spodoptera frugiperda (Lepidoptera, Noctuidae) from central Colombia. 7. Insect Bebav. 27:555–66
- 111. Scheidegger L, Niassy S, Midega C, Chiriboga X, Delabays N, et al. 2021. The role of *Desmodium intortum, Brachiaria* sp. and *Phaseolus vulgaris* in the management of fall armyworm *Spodoptera frugiperda* (J. E. Smith) in maize cropping systems in Africa. *Pest Manag. Sci.* 77:2350–57
- Schlum KA, Lamour K, de Bortoli CP, Banerjee R, Meagher R, et al. 2021. Whole genome comparisons reveal panmixia among fall armyworm (Spodoptera frugiperda) from diverse locations. BMC Genom. 22:179
- Schöfl G, Dill A, Heckel DG, Groot AT. 2011. Allochronic separation versus mate choice: nonrandom patterns of mating between fall armyworm host strains. Am. Nat. 177:470–85
- Schöfl G, Heckel DG, Groot AT. 2009. Time-shifted reproductive behaviours among fall armyworm (Noctuidae: Spodoptera frugiperda) host strains: evidence for differing modes of inheritance. J. Evol. Biol. 22:1447–59
- Shylesha AN, Jalali SK, Gupta A, Varshney R, Venkatesan T, et al. 2018. Studies on new invasive pest *Spodoptera frugiperda* (J. E. Smith) (Lepidoptera: Noctuidae) and its natural enemies. *J. Biol. Control* 32:145–51
- Sisay B, Simiyu J, Malusi P, Likhayo P, Mendesil E, et al. 2018. First report of the fall armyworm, Spodoptera frugiperda (Lepidoptera: Noctuidae), natural enemies from Africa. 7. Appl. Entomol. 142:800–4
- Sisay B, Tefera T, Wakgari M, Ayalew G, Mendesil E. 2019. The efficacy of selected synthetic insecticides and botanicals against fall armyworm, Spodoptera frugiperda, in maize. Insects 10:2

- 118. Smith JE, Abbot J. 1797. The Natural History of the Rarer Lepidopterous Insects of Georgia. Including Their Systematic Characters, the Particulars of the Several Metamorphoses, and the Plants on Which They Feed. Collected From the Observation of Mr. John Abbot, Many Years Resident in That Country. London: T. Bensley
- Song Y, Yang X, Zhang H, Zhang D, He W, et al. 2021. Interference competition and predation between invasive and native herbivores in maize. J. Pest Sci. 94:1053–63
- Sparks AN. 1980. Pheromones: potential for use in monitoring and managing populations of the fall armyworm. Fla. Entomol. 63:406–10
- Storer NP, Babcock JM, Schlenz M, Meade T, Thompson GD, et al. 2010. Discovery and characterization of field resistance to Bt maize: Spodoptera frugiperda (Lepidoptera: Noctuidae) in Puerto Rico. 7. Econ. Entomol. 103:1031–38
- 122. Sun X-x, Hu C-x, Jia H-r, Wu Q-l, Shen X-j, et al. 2021. Case study on the first immigration of fall armyworm, *Spodoptera frugiperda* invading into China. *7. Integr. Agric.* 20:664–72
- Tay WT, Behere GT, Batterham P, Heckel DG. 2010. Generation of microsatellite repeat families by RTE retrotransposons in lepidopteran genomes. BMC Evol. Biol. 10:144
- 124. Tay WT, Rane R, Padovan A, Walsh T, Elfekih S, et al. 2022. Global FAW population genomic signature supports complex introduction events across the Old World. Commun. Biol. 5:297
- 125. Tay WT, Rane RV, James W, Gordon KHJ, Downes S, et al. 2022. Resistance bioassays and allele characterisation inform analysis of Spodoptera frugiperda (Lepidoptera: Noctuidae) introduction pathways in Asia and Australia. J. Econ. Entomol. 115. In press
- Tefera T, Goftishu M, Ba M, Muniappan R. 2019. A Guide to Biological Control of Fall Armyworm in Africa Using Egg Parasitoids. Nairobi, Kenya: icipe
- 127. Tepa-Yotto GT, Meagher RL, Winsou JK, Dahoueto BTA, Tamò M, et al. 2022. Monitoring Spodoptera frugiperda in Benin: assessing the influence of trap type, pheromone blends, and habitat on pheromone trapping. Fla. Entomol. 105:71–78
- Tessnow AE, Gilligan TM, Burkness E, De Bortoli CP, Jurat-Fuentes JL, et al. 2021. Novel real-time PCR based assays for differentiating fall armyworm strains using four single nucleotide polymorphisms. Peer 79:e12195
- Tumlinson JH, Mitchell ER, Teal PEA, Heath RR, Mengelkoch LJ. 1986. Sex pheromone of fall army-worm, Spodoptera frugiperda (J.E. Smith)—identification of components critical to attraction in the field. 7. Chem. Ecol. 12:1909–26
- Unbehend M, Hänniger S, Meagher RL, Heckel DG, Groot AT. 2013. Pheromonal divergence between two strains of Spodoptera frugiperda. J. Chem. Ecol. 39:364

 –76
- 131. Unbehend M, Hänniger S, Vásquez GM, Juárez ML, Reisig D, et al. 2014. Geographic variation in sexual attraction of Spodoptera frugiperda corn- and rice-strain males to pheromone lures. PLOS ONE 9:11
- Varella AC, Menezes-Netto AC, Alonso JDD, Caixeta DF, Peterson RKD, Fernandes OA. 2015. Mortality dynamics of Spodoptera frugiperda (Lepidoptera: Noctuidae) immatures in maize. PLOS ONE 10-12
- Veenstra KH, Pashley DP, Ottea JA. 1995. Host-plant adaptation in fall armyworm host strains: comparison of food consumption, utilization, and detoxication enzyme activities. Ann. Entomol. Soc. Am. 88-80-91
- Vickery RA. 1929. Studies on the fall army worm in the Gulf coast district of Texas. Tech. Bull. 138, U. S. Dep. Agric., Washington, DC
- Vieira NF, Pomari-Fernandes A, Lemes AAF, Vacari AM, De Bortoli SA, de Freitas Bueno A. 2017. Cost of production of *Telenomus remus* (Hymenoptera: Platygastridae) grown in natural and alternative hosts. 7. Econ. Entomol. 110:2724–26
- 136. Vu TP. 2008. Insect pests of turf grass, biology, ecology and the control of Herpetogramma phaeoptralis (Guenée) in Hà Nội in Spring Summer 2008. MSc Thesis, Hà Nội Agric. Univ., Vietnam
- 137. Wang W, He P, Zhang Y, Liu T, Jing X, Zhang S. 2020. The population growth of *Spodoptera frugiperda* on six cash crop species and implications for its occurrence and damage potential in China. *Insects* 11:639
- Westbrook J, Fleischer S, Jairam S, Meagher R, Nagoshi R. 2019. Multigenerational migration of fall armyworm, a pest insect. *Ecosphere* 10(11):e02919

- Westbrook JK, Nagoshi RN, Meagher RL, Fleischer SJ, Jairam S. 2016. Modeling seasonal migration of fall armyworm moths. Int. 7. Biometeorol. 60:255–67
- Wiltshire EP. 1977. Middle East Lepidoptera, XXXVII: notes on the Spodoptera litura (F.)-group (Noctuidae—Trifinae). Proc. Br. Entomol. Nat. Hist. Soc. 10:92–96
- 141. Wu Q-L, He L-M, Shen X-J, Jiang Y-Y, Liu J, et al. 2019. Estimation of the potential infestation area of newly-invaded fall armyworm Spodoptera frugiperda in the Yangtze River Valley of China. Insects 10:298
- 142. Xiao H, Ye X, Xu H, Mei Y, Yang Y, et al. 2020. The genetic adaptations of fall armyworm *Spodoptera frugiperda* facilitated its rapid global dispersal and invasion. *Mol. Ecol. Resour.* 20:1050–68
- 143. Yainna S, Nègre N, Silvie PJ, Brévault T, Tay WT, et al. 2021. Geographic monitoring of insecticide resistance mutations in native and invasive populations of the fall armyworm. Insects 12:468
- 144. Yang F, Morsello S, Head GP, Sansone C, Huang F, et al. 2017. F2 screen, inheritance and cross-resistance of field-derived Vip3A resistance in *Spodoptera frugiperda* (Lepidoptera: Noctuidae) collected from Louisiana. USA. *Pest Manag. Sci.* 74:1769–78
- 145. Yathom S, Chen M, Tam S. 1981. The use of traps baited with species-specific pheromones for the detection of Spodoptera frugiperda. Phytoparasitica 9:221
- Young JR, McMillan WW. 1979. Differential feeding by two strains of fall armyworm larvae on carbaryl treated surfaces. J. Econ. Entomol. 72:202–3
- Yu S. 1991. Insecticide resistance in the fall armyworm, Spodoptera frugiperda (J. E. Smith). Pestic. Biochem. Physiol. 39:84–91
- 148. Zeng G, Zhi J-R, Zhang C-R, Zhang T, Ye J-Q, et al. 2021. Orius similis (Hemiptera: Anthocoridae): a promising candidate predator of Spodoptera frugiperda (Lepidoptera: Noctuidae). J. Econ. Entomol. 114:582–89
- Zhang L, Jin M, Zhang D, Jiang Y, Liu J, et al. 2019. Molecular identification of invasive fall armyworm Spodoptera frugiperda in Yunnan Province. Plant Prot. 45:19–24
- Zhang L, Liu B, Zheng W, Liu C, Zhang D, et al. 2020. Genetic structure and insecticide resistance characteristics of fall armyworm populations invading China. Mol. Ecol. Resour. 20:1682–96
- 151. Zimmerman EC. 1958. Insects of Hawaii. Honolulu, HI: Univ. Hawaii Press



Annual Review of Entomology

Contents

Volume 68, 2023

Complex and Beautiful: Unraveling the Intricate Communication Systems Among Plants and Insects *James H. Tumlinson**	1
Chemical Ecology of Floral Resources in Conservation Biological Control Stefano Colazza, Ezio Peri, and Antonino Cusumano	13
Management of Insect Pests with Bt Crops in the United States Aaron J. Gassmann and Dominic D. Reisig	31
Iron Homeostasis in Insects Maureen J. Gorman	51
Phoresy and Mites: More Than Just a Free Ride Owen D. Seeman and David Evans Walter	69
Postcopulatory Behavior of Tephritid Flies Diana Pérez-Staples and Solana Abraham	89
The Biology and Ecology of Parasitoid Wasps of Predatory Arthropods Minghui Fei, Rieta Gols, and Jeffrey A. Harvey	109
Dehydration Dynamics in Terrestrial Arthropods: From Water Sensing to Trophic Interactions Joshua B. Benoit, Kevin E. McCluney, Matthew J. DeGennaro, and Julian A.T. Dow	129
Biology and Management of the Spotted Lanternfly, <i>Lycorma delicatula</i> (Hemiptera: Fulgoridae), in the United States <i>Julie M. Urban and Heather Leach</i>	151
Historical and Contemporary Control Options Against Bed Bugs, Cimex spp. Stephen L. Doggett and Chow-Yang Lee	169
Functional Diversity of Vibrational Signaling Systems in Insects Meta Virant-Doberlet, Nataša Stritih-Peljhan, Alenka Žunič-Kosi, and Jernej Polajnar	191
Forest Insect Biosecurity: Processes, Patterns, Predictions, Pitfalls Helen F. Nahrung, Andrew M. Liebhold, Eckehard G. Brockerhoff, and Davide Rassati	211

Stingless Bee (Apidae: Apinae: Meliponini) Ecology David W. Roubik	231
Diapause in Univoltine and Semivoltine Life Cycles Hideharu Numata and Yoshinori Shintani	257
Early Monitoring of Forest Wood-Boring Pests with Remote Sensing Youqing Luo, Huaguo Huang, and Alain Roques	277
Spodoptera frugiperda: Ecology, Evolution, and Management Options of an Invasive Species Wee Tek Tay, Robert L. Meagher Jr., Cecilia Czepak, and Astrid T. Groot	299
Molecular Mechanisms of Winter Survival Nicholas M. Teets, Katie E. Marshall, and Julie A. Reynolds	319
Arthropod and Pathogen Damage on Fossil and Modern Plants: Exploring the Origins and Evolution of Herbivory on Land Conrad C. Labandeira and Torsten Wappler	341
The Resilience of Plant–Pollinator Networks Jordi Bascompte and Marten Scheffer	363
The Mechanisms of Silkworm Resistance to the Baculovirus and Antiviral Breeding Zhaoyang Hu, Feifei Zhu, and Keping Chen	381
Diversity, Form, and Postembryonic Development of Paleozoic Insects *Jakub Prokop, André Nel, and Michael S. Engel** **Long Company	401
Molecular Mechanisms Underlying Host Plant Specificity in Aphids Po-Yuan Shih, Akiko Sugio, and Jean-Christophe Simon	431
Adaptive Plasticity of Insect Eggs in Response to Environmental Challenges Monika Hilker; Hassan Salem, and Nina E. Fatouros	451

Errata

An online log of corrections to *Annual Review of Entomology* articles may be found at http://www.annualreviews.org/errata/ento